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Commentary: Identification of potential therapeutic compounds for Parkinson's disease using *Drosophila* and human cell models

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Parkinson's disease (PD) is the most common neurodegenerative movement disorder, affecting more than 1% of 55-year-old people and more than 3% of those older than 75. The best recognized pathological hallmark of PD is the loss of dopaminergic (DA) neurons in the substantia nigra, which leads to a decrease in dopamine levels in the striatum and causes movement impairment¹. Despite most PD cases are sporadic, several genes responsible for heritable forms of the disease have been described².³. Approximately 5-10% of patients suffer for a monogenic form of PD where dominant mutations in *SNCA*, *LRRK2* and *VP35* or recessive mutations in *PINK*, *DJ-1* and *Parkin* cause the disease with high penetrance². Functional studies of these genes have been essential to identify the mechanisms underlying the neuropathology of PD³. Interestingly, genome-wide association studies confirm that some of these genes are also affected in sporadic PD forms³.

Mutations in recessive PD genes like *PINK*, *DJ-1* and *Parkin* are relatively rare in the general PD population, but they are responsible of most cases of juvenile PD (mean onset age of ~39 years)². It has been shown that they regulate common pathways, being involved in mitochondrial quality control, protein degradation processes and oxidative stress response⁴. In our group, we are interested in *DJ-1*, which was first identified as a causative PD gene in consanguineous Dutch and Italian families that displayed early-onset PD⁵. Although it was initially described as an oncogene, subsequent studies have shown that it is also involved in mitochondrial homeostasis and defense against oxidative stress (OS)⁶. The DJ-1 protein is localized in the nucleus and the cytoplasm, but it is also present in mitochondria under OS conditions. Therefore, it has been proposed that DJ-1 is able to perform different functions depending on its subcellular localization⁷.

At present, PD is an incurable disease for which existing therapies are not sufficiently effective to stop or delay its progression. They are mainly based on a dopamine replacement strategy, capable of alleviating PD motor symptoms⁸. Common characteristics have been recently described in several neurodegenerative disorders such as increase in inflammation and OS levels, protein aggregation and ion homeostasis alterations⁹. For this reason, compounds with antioxidant, anti-inflammatory and neuroprotective activities are currently being considered as potential therapies for those disorders, including PD. Animal models of human diseases are commonly used in elucidating the biological pathways and environmental factors that influence their clinical presentation. In addition, they can be used in compound and small-molecule screening for the development of new therapeutic

strategies. In such a scenario, studies in *Drosophila* models have acquired a prominent position in providing insight into human biology and disease. Despite the apparent differences that do exist between *Drosophila* and humans, biological pathways and essential cellular processes are very similar in both organisms. This, together with its experimental tractability, makes *Drosophila* a very useful tool to identify new therapeutic compounds for human diseases ^{10,11}. In addition, chemical screens in a living animal such as *Drosophila* avoid common problems of traditional *in vitro* screens, in which lead compounds identified often fail to exert the expected effect in murine models due to suboptimal pharmacokinetics and pharmacodynamics ¹².

For several years, we have been working with a Drosophila model of familial PD based on inactivation of the DJ-1ß gene (ortholog of human DJ-1). Previous studies from our group demonstrated that DJ-1ß mutant flies exhibit motor defects, shortened lifespan, increased levels of reactive oxygen species (ROS) and other OS markers, and hypersensitivity to OS-inducing toxins¹³⁻¹⁵. Moreover, we also showed that some of these phenotypes could be suppressed with antioxidant treatments¹⁵. In an attempt to identify new potential drugs to treat PD, we recently analyzed the ability of several antioxidant, anti-inflammatory and neuroprotective compounds to suppress motor deficits of DJ-1ß mutants using a climbing assay¹⁶. This assay is a useful and easy method to evaluate locomotor ability in Drosophila, and is based on the negative geotactic behavior shown by this organism. It has been widely used to test the effectiveness of compounds to suppress motor deficits in *Drosophila* models of several neurodegenerative diseases, including familial PD based on mutations in SNCA¹⁷⁻²¹, LRRK2²²⁻²⁴, PINK1^{25,26} or parkin²⁷⁻³⁰. In our study, we selected 23 compounds already tested in humans and whose function or activity could be beneficial in a context of $DJ-1\beta$ deficiency. Additionally, their mechanisms of action were different from those of existing therapies used for PD treatment, and some of them were shown to be well-suited for other medical applications. Then, we performed a chemical screen to test the efficacy of those drugs to specifically suppress locomotor defects in DJ-1ß mutants. For doing so, model and control flies were cultured in medium supplemented with each compound during development plus five days after eclosion, and then were subjected to climbing assays. We found that some of them were beneficial for both control and DJ-1ß mutant flies, like celastrol, methylcobalamine, piracetam, metformin and acetylsalicylic acid, and others only for control flies, like curcumin. However, seven compounds were able to improve motor ability specifically in DJ-1ß mutant flies (dexrazoxane, pterostilbene, tocopherol, sodium phenylbutyrate, dalfampridine, methylene blue and minocycline), which were selected for subsequent analysis. At this point, we wondered how these compounds

could be exerting their beneficial effect in DJ-1ß mutants, and focused on their possible impact in ROS production and protein oxidative damage. Both human DJ-1 and DJ-1ß proteins protect against oxidative insults and are modified by oxidation^{6,31}. Consistently, DJ-1ß mutant flies displayed high levels of H₂O₂ (a component of the ROS pool) and of carbonylated proteins, when compared to control flies^{14,15}. Our results showed that while three of the seven candidate compounds were capable of reducing H₂O₂ levels (tocopherol, dalfampridine and minocycline), all of them but sodium phenylbutyrate also decreased the amount of carbonylated proteins. This is a very interesting finding because it suggests that this particular type of protein modification has a causative role in the locomotor defects of DJ-1ß mutants. Carbonyl groups are added to proteins when ROS levels are elevated, but also through metal-catalyzed oxidation³². In addition, non-oxidative mechanisms can also effectively increase carbonylation levels in proteins³³. Protein carbonylation is an irreversible posttranslational modification that generally leads to protein inactivation and degradation, therefore affecting cellular functions34. In such a scenario, our current interest is to identify the specific proteins affected by this damage, and probably inactivated, in PD model flies. This will help to further determine cellular processes underlying DJ-1ß inactivation and DJ-1-linked PD pathogenesis. Together, the results obtained in the Drosophila PD model indicate that the molecular mechanism by which the candidate compounds exert their beneficial effect are diverse, thus confirming the complexity of PD pathogenesis8.

Many studies performed in Drosophila models of human diseases have demonstrated that this organism can contribute to clinically relevant translational research in drug discovery¹². Therefore, to evaluate the translatability of the results obtained in flies we used undifferentiated human neuroblastoma SH-SY5Y cells, an in vitro model widely used in PD research³⁵. Several toxin-induced as well as genetic PD models have been obtained in these cells and used to test the therapeutic potential of compounds identified in other organisms^{24,36-38}. In our study, we generated DJ-1-deficient SH-SY5Y cells, in which the DJ-1 gene was silenced by RNAi¹⁶. Indeed, DJ-1 protein levels were reduced up to 90% in such cells. MTT assays showed that DJ-1-deficient cells were more susceptible to H₂O₂induced dose-dependent cytotoxicity than control cells, consistent with previous studies³⁹. Interestingly, we found that pretreatment of DJ-1 mutant cells with all candidate compounds but pterostilbene was able to reduce OS-induced cytotoxicity, thus supporting the therapeutic potential of the compounds identified in the Drosophila PD model. Previous studies have demonstrated that pterostilbene exerts its neuroprotective function by inhibiting microglial activation⁴⁰; hence, this compound is probably beneficial in DJ-1ß mutant flies (a whole animal model) through

microglial-like cells inactivation in the nervous system, despite being unable to suppress OS-induced cytotoxicity in DJ-1-deficient SH-SY5Y cells. In consequence, our study demonstrates that there is a good correlation between the results obtained in the Drosophila model and in DJ-1-deficient human cells, thus confirming the use of this organism in drug discovery for human diseases. It should be mentioned that all the candidate compounds identified in this work have been already tested in humans, and some of them have been used to treat patients with PD and other neurodegenerative disorders with different results. While some were able to alleviate symptoms in PD patients, others failed to provide improved clinical outcomes^{41,42}. Based on these data and considering the different mechanisms of action of the compounds identified, the development of a combinatorial therapy approach for PD targeting different cellular processes (not only OS) may result in additive or synergistic relief of pathology⁴¹.

Although *Drosophila* is not a substitute for preclinical testing in mammals, our study demonstrates that it can be an important component in drug discovery workflows when harnessed for efficient and high-throughput screenings⁴³. It also shows that the climbing assay is an easy method to identify compounds able to improve locomotor abilities in fly models of neurological disorders with movement impairments. This is especially relevant in the search of new effective therapies for PD because current treatments do not stop or delay the progression of the disease, and present problems in chronic treatments. As mentioned above, most PD cases are sporadic; however, several genes involved in rare familial forms of the disease appear to be affected in sporadic PD3. It has been shown that causative genes to PD like parkin or LRRK2 are misregulated in sporadic PD cases44,45. Regarding the DJ-1 gene, previous studies have demonstrated that the DI-1 protein is oxidatively modified, and probably inactivated, in sporadic PD cases⁴⁶. Besides, it has been recently suggested that α -Synuclein aggregates, which are the main components of Lewy bodies present in sporadic forms of PD, may recruit DJ-1 to form insoluble inclusions, thereby attenuating its antioxidant neuroprotective ability (see⁴⁷). Therefore, the results obtained in the *Drosophila* model of familial PD based on *DJ-1ß* inactivation, and validated in *DJ-*1-deficient human neuroblastoma cells, might be relevant for sporadic PD forms.

In conclusion, we hypothesize that any compound showing a rescue effect both in *Drosophila* and human cell PD models based on *DJ-1* deficiency would be likely to exert a beneficial effect in future clinical trials. However, further analyses will be required to confirm this assumption. Besides, we believe that the potential therapeutic compounds identified in our study could be useful to treat not only PD patients with mutations in the *DJ-1* gene but

also those affected by sporadic PD. The next step will be to perform high-throughput chemical screens using *DJ-1ß* mutant flies in an attempt to discover alternative and hopefully more effective PD treatments.

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